ANIMAL WORK

ADOPTIVE CELL TRANSFER

- Obtain Cell of Interest

* Depending on the level of purpose and specificity of the targeted cell population, choose your method for transfer cell obtainment such as magnetic beads or engineered T cells.
* This protocol uses the mashed spleen cells (splenocytes), and nylon sorted crude T cells

1. Obtain spleen and blood samples
2. Store blood at 4°C to obtain serum and mashed spleens to obtain splenocytes
3. Count the suspended cells
4. Prepare cells and cell numbers per transfer mouse (eg, 30M spleen cells /mouse, 10M T cells/mouse)
5. To use spleen T cells, remove red blood cells from the mashed spleen cells using ACK lysis buffer and perform nylon sorting.

- Transfer cells

6. Use proper injection/administration methods based on your purpose. Here, use tail vein injection.
7. Inject sterile PBS diluted cells (proper injection volume is 50 ~ 200 ul)

- Active tumor cell subcutaneous injection

8. Same day (Day 0) or one day later (Day 1), depending on your protocol, inject active tumor cells in the flank

- Observation of mice

9. You will see the death of mice around 1-2 weeks after tumor cell injection.
10. You may follow up to see the recruited T cells, or tumor sizes using IVIS imaging system.